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油红 O 染色试剂盒 (细胞专用)

货号: G1262

规格: 4×20mL/4×50mL

保存: 2-8℃, 避光保存, 有效期为 6 个月。

产品组成:

| VV - | | | | |
|--------------------|-----------------|--------|---------|-------------|
| 名称 | | 4×20mL | 4×50mL | 保存 |
| 试剂(A):油红O固定液 | | 20 mL | 50 mL | 室温, 避光 |
| 试剂(B):油红 | B1:油红O染色A液 | 12 mL | 30 mL | 2-8℃, 避光 |
| O染色液 | B2: 油红O染色B液 | 8 mL | 20 mL | 室温 |
| 按B1:B2=3:2比 | 例混合静置10min, 即为沟 | 由红O染色剂 | 友,不宜提前四 | 記制,过滤之后再使用。 |
| 试剂(C): Mayer苏木素染色液 | | 20mL | 50 mL | 2-8℃, 避光 |
| 试剂(D):油红O缓冲液 | | 20mL | 50 mL | 室温 |

产品介绍:

中性脂肪(Neutral fat)染色传统方法采用苏丹染料,最近发现偶氮染料油红 O 更适合脂肪的染色。常用染色方法有苏丹II、苏丹III、苏丹IV、苏丹黑 B、油红 O 法等。油红 O 是很强的脂溶剂和染脂剂,较易与甘油三脂结合呈小脂滴状,与磷脂结合力稍差,其染色原理一般认为是物理上的相似相溶作用或吸附作用,利用染料在细胞内脂质溶解率大于染液溶剂中溶解率来使脂肪染色。

油红 O 染色试剂盒(细胞专用)简称 ORO 染色液,可显示最小的脂滴,可优先为脂类从溶剂中吸附染料。标本不采用含有乙醇的固定液,如需要固定可采用 10%福尔马林)。脂肪阳性染色结果呈橘黄至红色,但具体颜色因脂质浓度而定。

自备材料:

60%异丙醇、蒸馏水

操作步骤: (仅供参考)

一、培养细胞

- 1. 移除细胞培养基,用PBS洗两次,加油红O固定液固定20-30min。
- 2. 弃去固定液,用蒸馏水洗2次,加入60%异丙醇浸洗20-30s。
- 3. 弃去60%异丙醇后加入新配制好的油红O染色液,浸染10-20min。
- 4. 弃去染色液,60%异丙醇漂洗20-30s至间质清晰。水洗2-5次,直到无多余染液。
- 5. 加入Mayer苏木素染色液,复染核1-2min。弃去染液后水洗2-5次,加入油红O缓冲液孵育 1min,弃去。
- 6. 加入蒸馏水覆盖细胞并在显微镜下观察。

二、细胞涂片

- 1. 制备新鲜骨髓、血液涂片,入油红O固定液固定10-15min,取出涂片,空气中晾干10-15min。
- 2. 入新配制好的油红O染色液,浸染15min。入60%异丙醇漂洗20-30s,流水冲洗,入蒸馏水稍微清洗。
- 3. 入Mayer苏木素染色液,复染核2min,入油红O缓冲液1min,镜下观察染色结果。

染色结果:

| 中性脂肪 | 橙红色或橘红色 | | |
|------|---------|--|--|
| 磷脂 | 粉红色 | | |
| 细胞核 | 蓝色 | | |

注意事项:

- 1. 油红O染色工作液不稳定, 易产生沉淀, 不宜提前配制。
- 2. Mayer苏木素染色液复染时间不能过长。
- 3. 染色结果不能长期保存,应尽快观察及照相。
- 4. 为了您的安全和健康,请穿实验服并戴一次性手套操作。

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Oil Red O Stain Kit, For Cultured Cells

Cat: G1262

Size:4×20mL/4×50mL

Storage: 2-8°C, avoid light, valid for 6 months.

Kit Components

| Reagent Reagent(A): ORO Fixative | | 4×20mL 20mL | 4×50mL 50 mL | Storage RT, avoid light |
|--|-------------------------------|-------------------|-----------------|----------------------------|
| | | | | |
| ORO Stain | B2: ORO Stain B | 8 mL | 20 mL | RT |
| Mix B1 with B2 | as the radio of 3:2 and place | for 10 min to for | rm ORO Stain, | filter it before use. |
| Reagent(C): Mayer Hematoxylin Solution | | 20mL | 50 mL | 2-8°C, avoid light |
| Reagent(D): ORO Buffer | | 20mL | 50 mL | RT |

Introduction

Neutral fat stains often use Sudan II, Sudan III, Sudan IV, Sudan black B, Oil red O and so on. Sudan dye is often used in traditional methods. Recently, azo dye oil red O is more suitable for dyeing fat. Oil Red O is a strong lipid solvent and dye. It is easy to bind to triglycerides in droplet shape, but has a slightly poor binding to phospholipids. The dyeing principle is generally regarded as a physical miscibility or adsorption, and the fat is dyed by miscibility. The solubility of dyes in frozen sections is higher than that in the original solvent, so when dyeing, the dyes are transferred from organic solvents to lipids and the fats are dyed.

Oil Red O Stain Kit, For Cultured Cells is shorten as ORO Stain and can show least fatty droplet. The sample should not fixed with ethanol (10% formalin if required). The positive staining result of fat is orange to red, but the specific color depended on the concentration of lipid.

Self Provided Materials

60% isopropanol, Distilled water

Protocol(for reference only)

For Cultured Cells

- 1. Remove the cell culture medium, wash it twice with PBS, and fix it with ORO Fixativ for 20-30min.
- Discard the ORO Fixative and wash it twice with distilled water.
- 3. Add 60% isopropanol and soak for 20-30s.
- 4. After discarding 60% isopropanol, add the newly prepared oil red O dyeing solution and dip it for 10-20min.
- 5. Discard the staining solution and rinse with 60% isopropanol for 20-30s until the stroma is clear. Wash with water 2-5 times until there is no excess dye.
- 6. Add Mayer hematoxylin staining solution and counterstain the nucleus for 1-2min. After discarding the dye solution, wash it with water for 2-5 times. Add oil red O buffer for 1min and discard it.
- 7. The cells were covered with distilled water and observed under a microscope.

For Cell Smear

- 1. Prepare fresh bone marrow and blood smear, then fix in ORO Fixative for 10-15 min.
- 2. Take out the smear and put it in the air for 10-15 min.
- 3. Soak the smear in ORO Stain and dye for 15 min.
- 4. Rinse with 60% isopropanol for 20-30s, rinse with running water, and wash slightly with distilled water. Re-dyeing the nucleus with Mayer Hematoxylin Solution for 1-2 min.
- 5. Soak in ORO Buffer for 1min and observed under a microscope..

Result

| Neutral Fat | Orange or Red | |
|--------------|---------------|--|
| Phospholipid | Pink Red | |
| Nucleus | Blue | |

Note

- 1. ORO Stain is not stable enough and easy to precipitate, so it is not suitable to prepare it in advance.
- 2. Mayer's Hematoxylin Solution should not stain for too long.
- 3. The dyeing results can not last for a long time, and should be viewed and photographed as soon as possible.
- 4. For your safety and health, please wear experimental clothes and disposable gloves.

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