V02

磷脂铁苏木素染色试剂盒(FeH 法)

货号：G3250
规格：3×100mL
保存：室温，避光保存，有效期 6 个月。

产品组成：

名称		3×100mL	保存
试剂(A): FeH 固定液		100mL	室温，避光
试剂(B): FeH 染色工作液	试剂(B1): FeH 苏木素染色液	25mL	室温，避光
	试剂(B2):FeH 缓冲液	75mL	室温，避光
将 B1 和 B2 按 1：3 的比例混匀即为 FeH 染色工作液，现配现用。			
试剂(C): FeH 分化液		100mL	室温

产品介绍：

磷脂(Phospholipid)是指含有磷酸的脂类，属于复合脂。磷脂是生物膜的成分，分为甘油磷脂和鞘磷脂两类。磷脂为两性分子，一端为亲水的含氮或磷的头，另一端为疏水（亲油）的长烃基链。Elleder 发现磷脂可通过三价铁苏木素显示出来，该法比 DAH 更简便、快速、敏感。但是需要使用丙酮对样本适当脱脂，而且会同时着色磷脂和细胞核。

自备材料：

氯仿、丙酮、甲醇、蒸馏水

操作步骤：(*仅供参考*)

1. 用氯仿甲醇溶液(氯仿：甲醇=1:1)浸泡 1 张切片 1h 作为阴性对照，实验切片用 2-8℃预冷的丙酮浸泡 5min。
2. 切片滴加 FeH 固定液固定 30min。蒸馏水冲洗 2 次，每次 3min。
3. 提前配制 FeH 染色工作液：B1 和 B2 按照 1：3 的比例混匀即为 FeH 染色工作液。
4. 组织切片滴加 FeH 染色工作液染色 7-10min。蒸馏水冲洗 2 次，每次 30s。
5. FeH 分化液加等量蒸馏水配成分化工作液，切片入分化工作液浸洗数次，每次 2-3s。 自来水冲洗。
6. 95%乙醇脱水，二甲苯透明，中性树脂封片。

染色结果：

磷脂	蓝色
细胞核	蓝色

注意事项：

1. 磷脂容易溶解，所以组织取出后应立即固定，否则难以着色。
2. 本染色液对冰冻切片的染色效果较好。
3. 染色后的标本务必避光保存，否则容易褪色。
4. 为了您的安全和健康，请穿实验服并戴一次性手套操作。

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Phospholipid-Fe Hematoxylin Stain Kit

Cat: G3250
Size: 3×100mL
Storage:RT, avoid light, valid for 6 months.

Kit Components

Reagent		3×100mL	Storage
Reagent(A): FeH Fixative		100mL	RT, avoid light
Reagent(B): FeH Stain Working Solution	Reagent(B1):Hematoxylin Solution	25mL	RT, avoid light
	Reagent(B2):FeH Buffer	75mL	RT, avoid light
Mix B1 and B2 in the ratio of 1:3 to prepare FeH Stain Working Solution.			
Reagent(C): FeH Differentiation		100mL	RT, avoid light

Introduction

Phospholipid is a kind of compound lipid containing phosphoric acid. Phospholipids are the components of biofilms, which are divided into glycerophospholipids and sphingolipids. Phospholipids are amphoteric molecules. One end is hydrophilic nitrogen or phosphorus containing head, and the other end is hydrophobic long alkyl chain. Elleder found that phospholipids can be displayed by ferritin, which is more simple, rapid and sensitive than Dah. However, acetone should be used to degrease the sample properly, and the phospholipid and nucleus will be stained at the same time.

Self Provided Materials

Chloroform, Acetone, Methanol, Distilled Water

Protocol(*for reference only*)

1. One slice was soaked in chloroform methanol solution (Chloroform: methanol = 1:1) for 1 h as negative control, and another slice was soaked in acetone precooled at 2-8 ℃ for 15 min.
2. Sections were fixed dropwise with FeH Fixative for 30 min. distilled water was rinsed twice for 3 min each.
3. Before dyeing, mix Reagent(B1) and Reagent(B2) in 1： 3 to prepare Reagent(B): FeH Stain Working Solution .
4. Tissue sections were stained dropwise with FeH Stain Working Solution for 7-10 min. Rinse with distilled water twice, 30s each time.
5. Dissolve the FeH Differentiation with distilled water 1:1 dilution, the slices were immersed in the diluted FeH differentiation solution for several times.Rinse with tap water.
6. Dehydrated with 95% ethanol, transparent with xylene and sealed with neutral resin.

Result

Phospholipid	Blue
Nucleus	Blue

Note

1. Phospholipids are easy to dissolve, so the tissue should be fixed immediately after removal, otherwise it is difficult to stain.
2. The results showed that the staining solution had good staining effect on frozen section.
3. The stained specimen must be kept away from light, otherwise it is easy to fade.
4. For your safety and health, please wear lab clothes and disposable gloves.