

## 切缘标记液（黑色）

货号：G4853

规格：100mL

保存：室温保存，有效期 1 年。

### 产品介绍：

切下组织之后暴露的切面称之为切缘，这一概念常用于肿瘤切除手术中。组织刚切下来时形态保留完好，可以较清晰的分辨出切缘的方位，但是当经过固定、逐级脱水直到制作成切片之后，分辨切缘的方位变得比较困难。然而在对手术切除的组织进行宏观检查时，通常需要永久的确定标本的方向，以便进一步进行大体检查和对切除边缘进行显微镜鉴定。

本品精选优质黑色颜料搭配成膜溶剂配制而成，经过本试剂标记后的组织在经过脱水和包埋之后仍然有足够厚度的颜色存留，着色牢固度高。本产品可以选择取新鲜组织直接涂抹标记，也可以将组织固定之后标记，选择将组织固定之后标记可以更好的保证组织结构稳定。

### 操作步骤：（仅供参考）

#### 无固定样本标记

1. 取材的新鲜组织用生理盐水洗去切面残留血液。
2. 用滤纸或纸巾吸去表面残余水分，必要时可用丙酮或 2%冰乙酸擦拭切面提高标记液粘附性。
3. 使用棉签蘸取适量标记液涂抹切缘，静置 3-5min 使染料充分黏附成膜。
4. 液氮或干冰乙醇混合浆速冻制备速冻切片。

#### 固定样本标记

1. 组织样本充分固定后取出，用滤纸或纸巾吸去表面残余水分，必要时可用丙酮或 2%冰乙酸擦拭切面提高标记液粘附性。
2. 使用棉签蘸取适量标记液涂抹或者蘸取在组织切缘，静置 3-5min 使染料充分黏附成膜。
3. 将标记后的组织放回固定液中固定 1h 以上。
4. 可以选择将组织继续固定或者直接脱水包埋。

### 注意事项：

1. 丙酮或 2%冰乙酸能增强标记液黏附性，但也会导致边缘轻度脱水变形，可根据样本状况酌情使用。
2. 标记颜料层的厚度应该适宜，保证有足够着色但是也不能太厚影响干燥速度。
3. 标记后的组织应尽量避免用镊子等硬物直接接触标记涂层。
4. 为了您的安全和健康，请穿实验服并戴一次性手套操作。

### 相关产品：

- G4851 切缘标记液（蓝色）
- G4852 切缘标记液（绿色）
- G4859 切缘标记液（三色）
- G2161 中性福尔马林固定液（10%）
- P1110 4%组织细胞固定液





## Tissue-Marking Dye, Black

**Cat:** G4853

**Size:** 100mL

**Storage:** RT, valid for 1 year.

### Introduction

The cut surface exposed after tissue removal is called the cutting edge, which is commonly used in tumor resection surgery. When the tissue is first cut off, its shape remains intact and the orientation of the cutting edge can be clearly distinguished. However, after fixation, gradual dehydration, and slicing, it becomes more difficult to distinguish the orientation of the cutting edge. However, when conducting macroscopic examination of the tissue removed by surgery, it is usually necessary to permanently determine the direction of the specimen for further macroscopic examination and microscopic identification of the resection edge.

This product is formulated with high-quality black pigments and film-forming solvents. After being labeled with this reagent, the tissue still retains sufficient thickness of color after dehydration and embedding, resulting in high color fastness. This product can choose to directly apply markings on fresh tissue, or mark after fixing the tissue. Choosing to mark after fixing the tissue can better ensure the stability of the organizational structure.

### Protocols(for reference only)

#### For unfixed sample

1. Wash the fresh tissue obtained with physiological saline to remove residual blood on the cut surface.
2. Use filter paper or tissue to absorb residual moisture on the surface, and if necessary, wipe the cut surface with acetone or 2% glacial acetic acid to improve the adhesion of the marking solution.
3. Use a cotton swab to dip an appropriate amount of Tissue-Marking Dye, Black onto the cutting edge, and let it stand for 3-5 min to fully adhere the dye into a film.
4. Liquid nitrogen or dry ice ethanol mixed slurry for rapid freezing to prepare frozen slices.

#### For fixed sample

1. After the tissue sample is fully fixed, remove it and use filter paper or tissue to absorb residual surface moisture. If necessary, acetone or 2% glacial acetic acid can be used to wipe the cut surface to improve the adhesion of the labeling solution.
2. Select the Tissue-Marking Dye, Black to apply or dip it onto the tissue cutting edge, and let it stand for 3-5 min to fully adhere the dye into a film.
3. Place the labeled tissue back into the fixative and fix for at least 1 h.
4. You can choose to continue fixing the tissue or directly dehydrate and embed it.

### Note

1. Acetone or 2% glacial acetic acid can enhance the adhesion of the labeling solution, but it can also cause slight dehydration and deformation of the edges. It can be used according to the sample condition.
2. The thickness of the marking pigment layer should be appropriate to ensure sufficient coloring, but it should not be too thick to affect the drying speed.
3. The labeled tissue should avoid direct contact with the labeled coating using forceps or other hard objects as much as possible.
4. For your safety and health, please wear laboratory clothes and disposable gloves to exercise.

