Coenzyme I NAD(H) Content Assay Kit

Note： It is necessary to predict 2-3 large difference samples before the formal determination.

Operation Equipment: Spectrophotometer

Cat No: BC0310

Size: 50T/24S

Product Composition:

Reagent name	Size	Preservation Condition
Acid Extract solution	Liquid 15 mL×1	2-8℃
Alkaline Extract solution	Liquid 15 mL×1	2-8℃
Reagent I	Liquid 20 mL×1	2-8℃
Reagent II	Liquid 6 mL×1	2-8℃
Reagent III	Liquid 16 mL×1	2-8℃
Reagent IV	Liquid 3 mL×1	-20℃
Reagent V	Liquid 40 mL×1	2-8℃
Reagent VI	Liquid 75 mL×1(Requird but not provided)	RT
NAD Standard	Powder×1	-20℃
NADH Standard	Powder×1	-20℃

Solution Preparation:

1. Reagent III: Need strict light protection.
2. Reagent VI: Take 72 mL of alcohol and 3 mL of distilled water, mix thoroughly.
3. NAD standard: Add 1.5 mL of distilled water before use to prepare as 2 μmol/mL, then dilute it to 1.25 nmol/mL NAD standard solution.
4. NADH standard: Add 1.4 mL of distilled water before use to prepare as 2 μmol/mL, then dilute it to 1.25 nmol/mL NADH standard solution.

Product Description：

Coenzyme I include both reduced and oxidized forms and plays a role in hydrogen transfer in biological oxidation. Oxidized coenzyme I also called nicotinamide adenine dinucleotide (NAD+) which is the coenzyme of dehydrogenase. It is an important role in glycolysis, gluconeogenesis, tricarboxylic acid cycle and respiratory chain. Intermediate product will transfer hydrogen to NAD make it become NADH (reduced coenzyme I). NADH acts as a carrier for hydrogen and synthesizes ATP by chemosmosis coupling in the respiratory chain. NADH has important physiological significance in the body. It is closely

related to substance metabolism, energy metabolism, anti-cell aging, anti-oxidation and the occurrence of some diseases. A decrease in coenzyme I levels in the body can lead to cell damage or death.

Extract the sample of NAD^+ and NADH with acidic and alkaline extract solution respectively. NADH reduces the oxidized Thiazolyl Blue Tetrazolium Blue (MTT) to form formazan by hydrogen transfer from

PMS, formazan has characteristic absorption at 570 nm. NAD could be reduced to NADH by alcohol dehydrogenase. Further, MTT reduction method was used to detect NAD^+.

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Required but Not Provided:

Spectrophotometer, water bath, desk centrifuge, transferpettor, 1 mL glass cuvette, mortar/ homogenizer, ice and distilled water.

Protocol

I. Extraction of NAD^+ and NADH:

1. Serum

Extraction of NAD^+: Add 0.5 mL of acid extract solution to 0.1 mL of serum, boiling 5 minutes(Wrap the sealing film to prevent bursting), ice bath after cooling. Centrifuge at 10000 ×g for 10 minutes at 4℃, take supernatant 200 μL and add equal volume alkaline extract solution. Mix thoroughly, centrifuge at 10000 ×g for 10 minutes at 4℃, take supernatant, preserve on ice for test.

Extraction of NADH: Add 0.5 mL of alkaline extract solution to 0.1 mL of serum, boiling 5 min(Wrap the sealing film to prevent bursting), ice bath after cooling. Centrifuge at 10000 ×g for 10 minutes at 4℃, take supernatant 200 μL and add equal volume acid extract solution. Mix thoroughly, centrifuge at 10000 ×g for 10 minutes at 4℃, take supernatant, preserve on ice for test.

2. Tissue

Extraction of NAD^+: Add 0.5 mL of acid extract solution to 0.1 g of tissue, grinding on ice, boiling 5 minutes(Wrap the sealing film to prevent bursting), ice bath after cooling. Centrifuge at 10000 ×g for 10 minutes at 4℃, take supernatant 200 μL and add equal volume alkaline extract solution. Mix thoroughly, centrifuge at 10000 ×g for 10 minutes at 4℃, take supernatant, preserve on ice for test.

Extraction of NADH: Add 0.5 mL of alkaline extract solution to 0.1 g of tissue, grinding on ice, boiling 5 minutes(Wrap the sealing film to prevent bursting), ice bath after cooling. Centrifuge at 10000 ×g for 10 minutes at 4℃, take supernatant 200 μL and add equal volume acid extract solution. Mix thoroughly, centrifuge at 10000 ×g for 10 minutes at 4℃, take supernatant, preserve on ice for test.

3. Cells or microorganism:

Extraction of NAD^+: Add 0.5 mL of acid extract solution to 5 million cells or germ, ultrasonic 1min(power 200W, ultrasonic 2s, interval 1s), boiling 5 minutes(Wrap the sealing film to prevent bursting), ice bath after cooling. Centrifuge at 10000 ×g for 10 minutes at 4℃, take supernatant 200 μL and add equal volume alkaline extract solution. Mix thoroughly, centrifuge at 10000 ×g for 10 minutes at 4℃, take supernatant, preserve on ice for test.

Extraction of NADH: Add 0.5 mL of alkaline extract solution to 5 million cells or germ, ultrasonic 1min (power 200W, ultrasonic 2s, interval 1s) boiling 5 minutes(Wrap the sealing film to prevent bursting), ice
bath after cooling. Centrifuge at 10000 ×g for 10 minutes at 4℃, take supernatant 200 μL and add equal
volume acid extract solution. Mix thoroughly, centrifuge at 10000 ×g for 10 minutes at 4℃, take supernatant, preserve on ice for test.

II. Procedure:

1.Preheat spectrophotometer for 30 minutes, adjust wavelength to 570 nm, set zero with distilled water.

2.Add reagents according to the following table.

Reagents	Contrast tube (μL)	Test tube (μL)	NAD or NADH	Blank tube

Supernatant	50	50	-	-
Standard	-	-	50	-
Distilled water				50
Reagent V	500	-	-	-

Reagent I	250	250	250	250
Reagent II	75	75	75	75
Reagent III	150	150	150	150
Reagent IV	35	35	35	35
Mix well and leave for 20 minutes at room temperature away from light				
Reagent V		500	500	500
Mix thoroughly, place for 5 minutes, centrifuge at 15000 rpm at 25°C for 15 minutes, discard supernatant.				
Reagent VI	1000	1000	1000	1000

Mix thoroughly, colorimetric at 570 nm, record the absorbance $\Delta A_T=A_T-A_C$, the standard tube of NAD, record $\Delta A_{S1}=A_{S1}-A_B$. The standard tube of NADH, record $\Delta A_{S2}=A_{S2}-A_B$. Blank tube just need test one to twice.

Calculation

The Content of NAD^+

1. Serum (plasma) sample

$NAD^+(nmol/mL)=\Delta A_T\div(\Delta A_{S1}\div C_S)\times V_E\div Vs=12.5\times\Delta A_T\div\Delta A_{S1}$

2. Tissue, germ or cells

1) Protein concentration

NAD^+ (nmol/mg prot)$=\Delta A_T\div(\Delta A_{S1}\div C_S)\times V_E\div(V_E\times Cpr)= 1.25\times\Delta A_T\div\Delta A_{S1}\div$ Cpr

2) Sample weight

NAD^+ (nmol/g)$=\Delta A_T\div(\Delta A_{S1}\div C_S)\times V_E\div W= 1.25\times\Delta A_T\div\Delta A_{S1}\div W$

3) Cells or germ

NAD^+ (nmol/10^4cell)$=\Delta A_T\div(\Delta A_{S1}\div C_S)\times V_E\div 500=0.0025\times\Delta A_T\div\Delta A_{S1}$

The Content of NADH

1. Serum (plasma) sample

NADH (nmol/mL) $=\Delta A_T\div(\Delta A_{S2}\div C_S)\times V_E\div V_{SE}==12.5\times\Delta A_T\div\Delta A_{S2}$

2. Tissue, germ or cells

1) Protein concentration

NADH (nmol/mg prot) $=\Delta A_T\div(\Delta A_{S2}\div C_S)\times V_E\div(V_S\times Cpr)= 1.25\times\Delta A_T\div\Delta A_{S2}\div$ Cpr

2) Sample weight

NADH (nmol/g) $=\Delta A_T\div(\Delta A_{S2}\div C_S)\times V_E\div W=1.25\times\Delta A_T\div\Delta A_{S2}\div W$

3) Cells or germ

NADH (nmol/10^4cell)$=\Delta A_T\div(\Delta A_{S2}\div C_S)\times V_E\div 500=0.0025\times\Delta A_T\div\Delta A_{S2}$

C_S：Concentration of NAD and NADH standard, 1.25 nmol/mL；

Cpr: Protein concentration, mg/mL;

V_E：Extract solution volume, 1 mL；

V_{SE}：Serum volume, 0.1 mL；

W：Sample weight, g；
500: 5 million cells.

Note：

1. Avoid light during operation. Do not mix Reagents I, II, III and then add, must be added separately.
2. Avoid light during reaction.
3. If the absorbance more than 1, measure the sample after diluting, multiply dilution times during equation.

Experimental examples：

1. Extraction of NAD^+: weigh about 0.1 g of lung, add 0.5 mL of Acid extract solution, grind in ice bath, boil for 5 min (cover tightly to prevent water loss), cool in ice bath, centrifuge at 4℃ and 10000g for 10 min, take 200 μL supernatant, add equal volume of Alkaline extract solution, mix well, After centrifugation at 4℃ and 10000g for 10 min, the supernatant is taken out, and then the operation is carried out according to the determination steps. The results showed that $\Delta A_T = A_T - A_C = 0.341-0.321 = 0.02$, $\Delta A_{S1} = A_{S1} - A_B = 1.045-0.246 = 0.799$, the content of NAD^+ is calculated according to the sample mass: NAD^+ (nmol/g mass) $= 1.25 \times \Delta A_T \div A_{S1} \div W = 1.25 \times 0.02 \div 0.799 \div 0.1 = 0.3129$ nmol/g mass.

Extraction of NADH: weigh about 0.1g lung, add 0.5ml Alkaline extract solution, grind in ice bath, boil for 5min (cover tightly to prevent moisture), cool in ice bath, centrifuge at 4℃ and 10000g for 10 min, take 200 μL supernatant, add equal volume of Acid extract solution, mix well, After centrifugation at 4℃ and 10000g for 10 min, the supernatant is taken out, and then the operation is carried out according to the determination steps. The results showed that $\Delta A_T = A_T - A_C = 0.2-0.136 = 0.064$, $\Delta A_{S2} = A_{S2} - A_B = 0.687-0.252 = 0.435$, the content of NADH is calculated according to the sample mass: NADH (nmol/g mass) $= 1.25 \times \Delta A_T \div A_{S2} \div W = 1.25 \times 0.064 \div 0.435 \div 0.1 = 1.839$ nmol/g mass.

2. Extraction of NAD^+: weigh about 0.1 mL of horse serum, add 0.5 mL of Acid extract solution, grind in ice bath, boil for 5 min (cover tightly to prevent water loss), cool in ice bath, centrifuge at 10000g and 4℃
for 10min, take 200 μL of supernatant, add equal volume of Alkaline extract solution, mix well, After centrifugation at 4℃ and 10000g for 10 min, the supernatant is taken out, and then the operation is carried out according to the determination steps. The results showed that $\Delta A_T = A_T - A_C = 0.091-0.061 = 0.03$, $\Delta A_{S1} = A_{S1} - A_B = 1.045-0.246 = 0.799$, NAD^+ content (nmol/mL) $= 12.5 \times \Delta A_T \div \Delta A_{S1} = 12.5 \times 0.03 \div 0.799 = 0.4693$ nmol/mL.

Extraction of NADH: weigh about 0.1 mL of horse serum, add 0.5 mL of Alkaline extract solution, grind in ice bath, boil for 5min (cover tightly to prevent moisture), cool in ice bath, centrifuge 10000g at 4 ℃ for 10min, take 200 μ l supernatant, add equal volume of Acid extract solution, mix well, After centrifugation at 4℃ and 10000g for 10 min, the supernatant is taken out, and then the operation is carried out according to the determination steps. The results showed that $\Delta AT = AT - AC = 0.126-0.1 = 0.026$, $\Delta A_{S2} = A_{S2} - A_B = 0.687-0.252 = 0.435$, the NADH content (nmol/mL) $= 12.5 \times \Delta A_T \div A_{S2} = 12.5 \times 0.026 \div 0.435 = 0.7471$ nmol/mL.

Recent Product Citation：

[1] Jiang Y, Cao S, Zhou B, Cao Q, Xu M, Sun T, Zhao X, Zhou Z, Wang Y. Hemocytes in blue

mussel Mytilus edulis adopt different energy supply modes to cope with different BDE-47 exposures. Sci Total Environ. 2023 Aug 10;885:163766. doi: 10.1016/j.scitotenv.2023.163766. Epub 2023 May 3. PMID: 37146804.

References:

[1] Ying W. NAD^+/NADH and $NADP^+$/NADPH in cellular functions and cell death: regulation and biological consequences[J]. Antioxidants & redox signaling, 2008, 10(2): 179-206.

[2] Gibon Y, Larher F. Cycling assay for nicotinamide adenine dinucleotides: NaCl precipitation and ethanol solubilization of the reduced tetrazolium[J]. Analytical biochemistry, 1997, 251(2): 153-157.

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