

Thioredoxin Reductase (TrxR) Activity Assay Kit

Note: Take two or three different samples for prediction before test.

Operation Equipment: Spectrophotometer/Microplate Reader

Cat No: BC1155 Size:100T/96S

Product Composition: Before use, please carefully check whether the volume of the reagent is consistent with the volume in the bottle. If you have any questions, please contact Solarbio staff in time.

Reagent name	Size	Preservation Condition
Reagent I	Liquid 125 mL×1	2-8°C
Reagent II	Liquid 3 mL×1	2-8°C
Reagent III	Powder×2	-20°C
Reagent IV	Liquid 30 μL×1	-20°C

Solution Preparation:

- 1. Reagent III: The reagent was placed in a glass bottle, and before use, one bottle was dissolved in 1.667 mL distilled water, and the reagent was stored at -20°C for 2 weeks.
- 2. Reagent IV: Small volume, please centrifuge before use. Before clinical use, the reagent was diluted 10 times with anhydrous ethanol four times according to the number of samples.

Product Description:

TrxR is a NADPH-dependent dimer selenase and includes FAD structure domain. TrxR belongs to pyridine nucleotide-disulfide REDOX enzyme, and form thioredoxin system with thioredoxin and NADPH. The activity of TrxR is similar with GR. TrxR could catalyzes GSSG reduce to GSH, which is the key enzyme in glutathione REDOX cycle.

TrxR catalyzes NADPH to reduceDTNB form TNB and NADP⁺, TNB has a absorbance at 412 nm, but reduced glutathione reacts with DTNB to form TNB, so the 2-Vinylpyridine in this kit can inhibit reduced glutathione in sample, the activity of TrxR can be calculated by detecting increase rate of TNB at 412 nm.

Reagents and Equipment Required but Not Provided:

Spectrophotometer/microplate reader, ultra-micro glass cuvette/96 well flat-bottom plate,water bath, low temperature centrifuge, mortar/homogenizer, adjustable pipette,distilled water.

Procedure

I. Sample preparation:

1. Tissue:

Add 1 mL of Reagent I into 0.1 g of tissue, fully grinding on ice. Centrifuge at 10000 rpm for 10 minutes at 4°C and take the supernatant on ice for test. Before test, mix supernatant and Reagent IV at a



ratio of 50:1(add 2 μ L of Reagent IV to 100 μ L of supernatant), water bath at 37°C for 30 minutes, then keep on ice for test.

2. Bacteria/cell:

Suggested 5 million with 1 mL of Reagent I, Splitting bacteria and cell with ultrasonic(ice bath, power 300W, work time 3 s, interval 7 s, for 3 minutes), centrifuge at 10000 rpm and 4°C for 10 minutes. The supernatant on ice is used for test. before test, mix supernatant and Reagent IV at a ratio of 50:1(add 2 μ L of Reagent IV to 100 μ L of supernatant), water bath at 37°C for 30 minutes, then keep on ice for test.

III. Procedure:

- 1. Preheat spectrophotometer/microplate reader for 30 minutes, adjust the wavelength to 412 nm, set the zero with distilled water.
- 2. Preheat reagent1 at 37°C(mammal), 25°C(other) in water bath for 30 minutes.
- 3. Blank tube: take a micro glass cuvette/96 well flat-bottom plate, add 20 μ Lof Reagent II, 20 μ Lof Reagent III, 160 μ Lof Reagent I, mix them quickly, and then measure the absorbance at 412nm for 10s. Take out the absorbance at 412 nm in a 37°C water bath for 5min and record it as A1 and A2. Calculate $\triangle A_B = A2-A1$.
- 4. Measuring tube: take a micro glass cuvette/96 well flat-bottom plate, add 20 μ Lof Reagent II, 20 μ Lof Reagent III, 140 μ Lof Reagent I, 20 μ Lof supernatant, mix it quickly, and then measure the absorbance at 412nm for 10s, take out the absorbance at 412 nm quickly in a 37°C water bath for 5min, and record it as A3 and A4. \triangle A_T= A4-A3.

III. Calculation:

A. micro glass cuvette:

1. Protein concentration:

Unit definition: One unit of enzyme activity is defined as the amount of enzymecatalyzes the production of 1 µmol of TNB at 37°C(mammal), 25°C(other) per minuteevery mg of protein.

 $TrxR(U/mgprot) = [\Delta A(T) - \Delta A(B)] \div (\epsilon \times d) \times 109 \times Vrv \div (Vs \times Cpr) \div T = 147 \times [\Delta A(T) - \Delta A(B)] \div Cpr$

2. Sample weight:

Unit definition: One unit of enzyme activity is defined as the amount of enzymecatalyzes the production of 1 µmol of TNB at 37°C(mammal), 25°C(other) per minuteevery gram of sample.

 $TrxR(U/g) = [\Delta A(T) - \Delta A(B)] \div (\epsilon \times d) \times 10^9 \times Vrv \div (Vs \div Vsv \times W) \div T = 147 \times [\Delta A(T) - \Delta A(B)] \div W$

3. Cells:

Unit definition: One unit of enzyme activity is defined as the amount of enzyme catalyzes the production of 1 µmol of TNB at 37°C(mammal), 25°C(other) per minute every 10⁴ cell.

 $TrxR(U/10^{4}cell) = [\Delta A(T) - \Delta A(B)] \div (\epsilon \times d) \times 10^{9} \times Vrv \div (N \div Vsv \times Vs) \div T = 147 \times [\Delta A(T) - \Delta A(B)] \div N$

ε: TNB molar extinction coefficient, 1.36×10⁴ L/mol/cm;

d: Light path of cuvette, 1cm;

Vrv: Total reaction volume, 200 μ L=2×10⁻⁴ L;

Vs: Supernatant volume (mL), 0.02 mL;



Cpr: Sample protein concentration (mg/mL); need to detect separately, suggest use PC0020, BCA Protein Assay Kit;

T: Reaction time (min), 5 minutes;

W: Sample weight(g);

Vsv: Extract solution volume, 1 mL;

N: Amount of cells, 10⁴.

B. 96 well flat-bottom plate

1. Protein concentration:

Unit definition: One unit of enzyme activity is defined as the amount of enzymecatalyzes the production of 1 µmol of TNB at 37°C(mammal), 25°C(other) per minute every mg of protein.

TrxR (U/mg prot) =[
$$\Delta$$
A(T)- Δ A(B)]÷(ϵ ×d)×10⁹×Vrv÷(Vs×Cpr)÷T
= 245×[Δ A(T)- Δ A(B)]÷Cpr

2. Sample weight:

Unit definition: One unit of enzyme activity is defined as the amount of enzymecatalyzes the production of 1 µmol of TNB at 37°C(mammal), 25°C(other) per minute every gram of sample.

$$TrxR(U/g) = [\Delta A(T)-\Delta A(B)] \div (\epsilon \times d) \times 10^{9} \times Vrv \div (Vs \div Vsv \times W) \div T$$
$$= 245 \times [\Delta A(T)-\Delta A(B)] \div W$$

3. Cells:

Unit definition: One unit of enzyme activity is defined as the amount of enzyme catalyzes the production of 1µmol of TNB at 37°C(mammal), 25°C(other) per minute every 10⁴ cell.

$$\begin{aligned} TrxR(U/10^{4}cell) &= [\Delta A(T)-\Delta A(B)] \div (\epsilon \times d) \times 10^{9} \times Vrv \div (N \div Vsv \times Vs) \div T \\ &= 245 \times [\Delta A(T)-\Delta A(B)] \div N \end{aligned}$$

ε: TNB molar extinction coefficient at 412 nm,1.36×10⁴ L/mol/cm;

d: Light path of cuvette, 0.6 cm;

Vrv: Total reaction volume, 200 μ L=2×10⁻⁴ L;

Vs: Supernatant volume (mL), 0.02 mL;

Cpr: Sample protein concentration (mg/mL); need to detect separately, suggest use PC0020, BCA Protein Assay Kit;

T: Reaction time (min), 5 minutes;

W: Sample weight(g);

Vsv: Extraction volume, 1 mL;

N: Amount of cells, 10⁴.

Note:

1. Dilute 5 times with distilled water when detecting mammalian tissue and blood samples, detect quickly as soon as possible.

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2. Because the extract solution contains a certain concentration of protein (about 0.1 mg/mL), the protein content of the extract solution itself needs to be subtracted when determining the protein concentration of the sample.

Experimental instances:

1. Take 0.1g of Chinese rose petals, add 1mL of extract solution, fully grinding on ice. Centrifuge at 10000 rpm for 10 minutes at 4°C, take the supernatant and place it on ice for test according to the measured steps. Calculate $\Delta A_T = A4 - A3 = 0.8600 - 0.8177 = 0.0423$, $\Delta A_B = A2 - A1 = 0.0792 - 0.0727 = 0.0065$, calculate the enzyme activity according to sample weight: TrxR (U/g weight) = 147 × ($\Delta A_T - \Delta A_B$) $\div W = 5.26$ U/g weight.

2. Take 0.1g of liver, add 1mL of extract solution, fully grinding on ice. Centrifuge at 10000 rpm for 10 minutes at 4°C, take the supernatant and place it on ice for test according to the measured steps. Calculate $\Delta A_T = A4 - A3 = 0.8538 - 0.2102 = 0.6436$, $\Delta A_B = A2 - A1 = 0.0792 - 0.0727 = 0.0065$, calculate the enzyme activity according to sample weight:

TrxR (U/g weight) =147× (ΔA_T - ΔA_B) ÷W×4 (dilution ridio) =3746.148 U/g weight.

Recent Product citations

- [1] Li B, Li D, Jing W, et al. Biogenic selenium and its hepatoprotective activity[J]. Scientific reports, 2017, 7(1): 1-11.
- [2] Zhang L, Fan J, He J, et al. Regulation of ROS–NF-κB axis by tuna backbone derived peptide ameliorates inflammation in necrotizing enterocolitis[J]. Journal of cellular physiology, 2019, 234(8): 14330-14338.

Related products:

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