

β -amylase(β -AL) Activity Assay Kit

Note: It is necessary to predict 2-3 large difference samples before the formal determination.

Detection instrument: Spectrophotometer/microplate reader

Cat No: BC2045

Size: 100T/48S

Components:

Reagent I: Liquid 35 mL \times 1. Store at room temperature. If yellow crystal is precipitated, heated moderately to dissolve before use.

Reagent II: Liquid 10 mL \times 2. Store at 2-8°C. Each Reagent II is added to each Reagent III. The solution is placed in room temperature water, heat with frequent agitation and boil to completely dissolve the powder. It could be stored at 2-8°C for four weeks.

Reagent III: Powder \times 2. Store at 2-8°C.

Standard: Powder \times 1. Store at 2-8°C. 10 mg anhydrous glucose. Add 1 mL of distilled water to form 10 mg/mL glucose standard solution when the solution will be used. It could be stored at 2-8°C for two weeks.

Product Description:

Amylase is responsible for hydrolyzing starch, including α -amylase and β -amylase. β -amylase (EC 3.2.1.2) cuts α -1, 4 glycoside bonds from the non-reducing end of starch to produce glucose, maltose, maltose, dextrin and other reducing sugars.

Reducing sugar reduced 3,5-dinitrosalicylic acid to form brown red substance. α -amylase is acid-resistant and β -amylase is heat-resistant. According to the above characteristics, the activity of another amylase can be measured by passivating one of them.

Required material:

Spectrophotometer/microplate reader, thermostat water bath, centrifuge, transferpeltor, micro glass cuvette/96 well flat-bottom plate, mortar/homogenizer, distilled water.

Procedure:

I. Sample Extraction:

It is suggested that when weigh about 0.1 g of sample, add 0.8 mL of distilled water. After homogenize, place the extract at room temperature and extract for 15 minutes. Shake once every 5 minutes to fully extracted. Centrifuge at 6000 \times g for 10 minutes at room temperature. Take the supernatant and add distilled water to constant volume to 10 mL, shake well, that is the original amylase solution.

Take 1 mL of the above-mentioned amylase stock solution, add 4 mL of distilled water, shake well, it is the amylase diluent, which is used for the determination of the total activity of (α + β) amylase.

II. Determination procedure:

1 Preheat the spectrophotometer for 30 minutes, adjust wavelength to 540 nm, and set spectrophotometer counter to zero with distilled water.

2 Standard working solution: dilute the glucose standard solution with distilled water to 0.5, 0.25, 0.125,

0.0625, 0.03125, 0.015625 and 0.0078 mg/mL.

3 Add 75 μ L of amylase stock solution and diluted amylase solution into two EP tubes respectively, and use them as the contrast tube of α -amylase contrast tube and the contrast tube of β -amylase respectively after boiling for 5 minutes.

4 Measurement operation table:

Reagent (μ L)	Measured of α -amylase activity		Measured of total amylase activity		Measured of standard curve	
	Contrast tube (C)	Test tube (T)	Contrast tube (C)	Test tube (T)	Standard tube (S)	Blank tube (B)
Amylase stoste	75 (boiling)	75	-	-	-	-
Distilled water	-	-	-	-	-	75
Standard solution	-	-	-	-	75	-
Incubate in 70°C water bath for 15 minutes, cooling.						
Diluted amylase solution	-	-	75 (boiling)	75	-	-
Reagent II	-	75	-	75	-	-
Incubate in 40°C thermostat water bath for 5 minutes.						
Reagent I	150	150	150	150	150	150
Reagent II	75	-	75	-	75	75

Mix well, boiling water bath for 10 minutes, then add 200 μ L to micro glass cuvette/96-well plate, measure the absorbance at 540 nm. recorded as A_1, A_2, A_3, A_4, A_5 and A_6 respectively from left to right. $\Delta A_\alpha = A(2) - A(1)$, $\Delta A_{Total} = A(4) - A(3)$, $\Delta A_{Standard} = A(5) - A(6)$. Each test tube should be provided with one contrast tube. Standard curve and blank tube only need to be measured once or twice.

III. Calculation:

1、 Create standard curve

Using the concentration of standard solution as x axis and $\Delta A_{Standard}$ as y axis create standard curve, obtain equation $y = kx + b$. Put ΔA_α into the equation and obtain the x_1 (mg/mL), Put ΔA_{Total} into the equation and obtain the x_2 (mg/mL).

2、 Calculation of α -amylase activity

(1) Calculation according to sample quality

Unit definition: One unit of enzyme activity is defined as the amount of enzyme catalyzes the generation of 1 mg of reducing sugar in the reaction system per minute every g sample.

$$\alpha\text{-amylase activity (U/g fresh weight)} = x_1 \times V_S \div (W \times V_S \div V_{ST}) \div T = 2 \times x_1 \div W$$

(2) Calculation according to protein content

Unit definition: One unit of enzyme activity is defined as the amount of enzyme catalyzes the generation of 1 mg of reducing sugar in the reaction system per minute every mg protein.

$$\alpha\text{-amylase activity (U/mg prot)} = x_1 \times V_S \div (V_S \times C_{pr}) \div T = 0.2 \times x_1 \div C_{pr}$$

3、 Calculation of total amylase activity

(1) Calculation according to sample quality

Unit definition: One unit of enzyme activity is defined as the amount of enzyme catalyzes the

generation of 1 mg of reducing sugar in the reaction system per minute every g sample.

$$\text{Total amylase (U/g fresh weight)} = 5 \times x_2 \times V_S \div (W \times V_S \div V_{ST}) \div T = 10 \times x_2 \div W$$

(2) Calculation according to protein content

Unit definition: One unit of enzyme activity is defined as the amount of enzyme catalyzes the generation of 1 mg of reducing sugar in the reaction system per minute every mg protein.

$$\text{Total amylase (U/mg prot)} = 5 \times x_2 \times V_S \div (V_S \times C_{pr}) \div T = x_2 \div C_{pr}$$

4、 Calculation of β -amylase activity

(1) Calculation according to sample quality

Unit definition: One unit of enzyme activity is defined as the amount of enzyme catalyzes the generation of 1 mg of reducing sugar in the reaction system per minute every g sample.

$$\begin{aligned} \beta\text{-amylase activity (U/g fresh weight)} &= \text{The activity of total amylase} - \alpha\text{-amylase activity} \\ &= (10 \times x_2 \div W) - (2 \times x_1 \div W) = (10 \times x_2 - 2 \times x_1) \div W \end{aligned}$$

(2) Calculation according to protein content

Unit definition: One unit of enzyme activity is defined as the amount of enzyme catalyzes the generation of 1 mg of reducing sugar in the reaction system per minute every mg protein.

$$\begin{aligned} \beta\text{-amylase activity (U/mg prot)} &= \text{The activity of total amylase} - \alpha\text{-amylase activity} \\ &= (x_2 \div C_{pr}) - (0.2 \times x_1 \div C_{pr}) \end{aligned}$$

5: Dilution ratio of total amylase;

V_S : The volume of sample added to reaction system, 0.075 mL;

V_{ST} : Total volume of extract, 10 mL;

C_{pr} : Sample protein concentration, mg/mL;

T: reaction time, 5 min;

W: Sample weight, g.

Note:

When the measured absorbance value is greater than 1.5, the sample can be appropriately diluted for determination. If the absorbance value is too small, diluted amylase solution or amylase stock solution can be concentrated.

References:

[1] Dzedzoave N T, Graffham A J, Westby A, et al. Influence of variety and growth environment on β -amylase activity of flour from sweet potato (*Ipomea batatas*)[J]. Food control, 2010, 21(2): 162-165.

Related Products:

- BC0700/BC0705 Glutamate Synthase(GOGAT) Activity Assay Kit
- BC0610/BC0615 α -amylase Activity Assay Kit
- BC0430/BC0435 ADPG Pyrophosphorylase(AGP) Activity Assay Kit