

L-Lactic Acid(L-LA) Content Assay Kit

Note: The reagents have been changed, so please be aware of and follow this instruction strictly.

Operation Equipment: Spectrophotometer/ Microplate reader

Cat No: BC2235 **Size:** 100T/48S

Components:

Extract solution I: Liquid 60 mL×1 bottle, store at 2-8°C; Extract solution II: Liquid 10 mL×1 bottle, store at 2-8°C;

Reagent I: Liquid 6 mL×1 bottle, store at 2-8°C;

Reagent II: Liquid 20 μL×1 bottle, store at 2-8°C. Accordance ratio Reagent II: Distilled water=10 μL: 450 μL. Prepare when it will be used.

Reagent III: Liquid 8 mL×1 bottle, store at 2-8°C;

Reagent IV: Powder×1 bottle, store at -20°C; Before use, add 3 mL distilled water to each bottle and mix well. After packaging, store at -20°C for 4 weeks to avoid repeated freeze-thaw.

Reagent V: Liquid 2 mL×1 bottle, store at 2-8°C;

Standard: Powder×1, store at 2-8°C; Add 1.04 mL distilled water to 100 µmol/mL before use. It can be stored at 2-8°C for 4 weeks.

Description:

L (+)-Lactic acid is an important intermediate product in biological metabolism. It is closely related to glucose metabolism, lipid metabolism, protein metabolism and intracellular energy metabolism. Lactic content is an important indicator for assessing carbohydrate metabolism and aerobic metabolism. L (+)-Lactic produces pyruvic acid under the action of lactate dehydrogenase, and NAD+ is reduced to produce NADH and H+. H+ is transferred to PMS to produces PMSH2 and PMSH2 reduce MTT to form purple substance, and has a characteristic absorption peak at 570 nm.

Required but not provided:

Scale, mortar/homogenizer, centrifuge, spectrophotometer/microplate reader, micro glass cuvette/ 96 well flat-bottom plate, constant temperature water bath, ethanol, distilled water.

Procedure:

I. Sample Preparation.

1. Tissue:

Accordance ratio weight(g): Extract solution I(mL)=1: 5~10. (Suggested 0.1g tissue with 1mL Extract solution I). Homogenate on ice bath. 12000 g centrifuge for 10 min at 4°C. Add 0.15mL Extract solution II slowly to 0.8 mL supernatant. **Blend slowly until no bubbles.** Then 12000 g centrifuge for 10 min at 4°C. Supernatant is for test.



2. Cells:

Accordance ratio cell amount (10^6): Extract solution I(mL)= $5\sim10:1$. (Suggested 5×10^6 cells with 1mL Extract solution I). Breaking cells (300W, work time 3s, interval 7s for 3 min) by ultrasonic on ice bath. 12000 g centrifuge for 10 min at 4°C. Add 0.15 mL Extract solution II to 0.8mL supernatant slowly. **Blend slowly until no bubbles**. 12000 g centrifuge for 10 min at 4°C. Supernatant is used for test.

3. Serum (plasma) sample:

Add 1 mL Extract solution I to 100 μL serum(plasma). 12000 g centrifuge for 10 min at 4°C. Add 0.15 mL Extract solution II to 0.8 mL supernatant slowly. **Blend slowly until no bubbles.** Centrifuge for 10 min at 12000 g. Supernatant is used for test.

II. Determination procedure.

- 1. Preheat spectrophotometer/microplate reader for 30 min, adjust wavelength to 570 nm, spectrophotometer set zero with ethanol.
- 2. Standard working solution: $100\mu\text{mol/mL}$ standard was diluted with distilled water to be 2.5, 1.25, 0.625, 0.3125, 0.15625, $0.078\mu\text{mol/mL}$ for test.
- 3. Add reagents according to the following table.

	C		
Test tube (T)	Control tube (C)	Standard tube (S)	Blank tube (B)
10	10		
		10	Jar Photo
-	0 10	-	10
40	40	40	40
10	JENO -	10	10
20	20	20	20
oroughly in centri	ifuge tube, react 20	min at 37°C water bat	h.
6	6	6	6
60	60	60	60
rifuge tube. Avoid	ling light react 20 m	nin at 37°C, 10000 rpr	n centrifuge for
e supernatant and	l retain sediment.		CO JUNE
200	200	200	200
ents, and deter	mine absorbance	at 570 nm, record	A_T , A_C , A_S , A_B ,
$\Lambda_{ m B.}$.0i0s	
	10 40 10 20 broughly in centre 6 60 fifuge tube. Avoid a supernatant and 200 ents, and deter	10 10 - 10 40 40 10 - 20 20 20 broughly in centrifuge tube, react 20 6 6 6 60 60 iffuge tube. Avoiding light react 20 me supernatant and retain sediment. 200 200 ents, and determine absorbance and retain sediment.	10 10 - 10 - 10 - 10 - 10 - 10 20 20 20 20 20 20 20 20 20 6 6 6 60 60 60 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 20 60 200 200 200 200 200 200 ents, and determine absorbance at 570 nm, record 570 nm, record

III. Calculation.

1. Drawing of standard curve.

Standard solution concentration as x axis and its corresponding absorption value (ΔAs) as y axis, the standard equation is y=kx+b. Bring ΔA_T into the formula to get x (μ mol/mL).

2. Calculation of Lactate content.

A. Protein concentration:

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L-LA (μ mol/mg prot)= $x \times Vs \div (Vs \times Cpr)$ = $x \div Cpr$.

B. Sample weight

L-LA (μ mol/g weight)= $x \times (Vsp+V_{II}) \div (W \times Vsp \div V_{I}) = 1.1875 \times x \div W$.

C. Cell amount

L-LA (μ mol/10⁶ cell)= $x\times(Vsp+V_I)$ ÷($N\times Vsp\div V_I$)= 1.1875× $x\div N$.

D. Liquid volume

L-LA (μ mol/mL)= $x\times(Vsp+V_{II})$ ÷ [$V_L\times Vsp$ ÷(V_I+V_L)]= 13.0625×x

Vs: Sample volume, 0.01 mL

W: Sample weight, g

Cpr: Sample protein concentration, mg/mL(Protein concentration needs to be Self- determined)

Vsp: Supernatant volume, 0.8 mL

V_{II}: Extract solution II, 0.15 mL

V_I: Extract solution I, 1 mL

N: Number of cells, in millions.

V_L: Liquid sample volume, 0.1 mL

Note:

- 1. If the absorbance value exceeding 1.5 or ΔA >1.2, please dilute the sample volume properly and multiply the dilution multiple in the formula.
- 2. Extract I contains a protein precipitant and therefore the supernatant cannot be used for protein concentration determination. If protein content is to be determined, a separate sample is required.

Experimental example:

1. Take 0.1g of rabbit heart, add 1 mL of Extract solution 1, grind and centrifuge, take 0.8 mL of supernatant and add 0.15 ml of Extract solution 2, centrifuge supernatant and dilute 5 times, then operate according to the determination steps, use 96 well plate to measure and calculate $\Delta AT=AT-AC=0.591-0.069=0.522$, according to the standard curve y=0.412x-0.0214, x=1.319, calculate the content according to the sample mass.

La content (μ mol/g mass) = 1.1875×x ÷ W × dilution ratio =1.1875 ×1.319 ÷ 0.1 × 5 = 78.32 μ mol/g mass.

2. Take 100 μ L of mouse serum, add 1 mL of Extract solution 1, take 0.8 mL of supernatant and then add 0.15 mL of Extract solution 2, centrifugate the supernatant, and then operate according to the determination steps, use 96 well plate to measure and calculate $\Delta A_T = A_T - A_C = 0.572 - 0.211$



=0.361, according to the standard curve y = 0.412x-0.0214, x = 0.928, calculate the content according to the liquid volume

La content (μ mol/mL) = 13.0625 × x = 13.0625 × 0.928 = 12.122 μ mol/mL.

Recent Product Citations:

[1] Meixi Peng, Dan Yang, Yixuan Hou, et al. Intracellular citrate accumulation by oxidized

ATM-mediated metabolism reprogramming via PFKP and CS enhances hypoxic breast cancer cell invasion and metastasis. Cell Death and Disease. March 2019;(IF5.959)

- [2] Xiaojin Luo, Weihua Shi, Haoming Yu, et al. Wearable Carbon Nanotube-Based BioSensors on Gloves for Lactate. Sensors. October 2018;(IF3.031)
- [3] Zhou F, Du J, Wang J. Albendazole inhibits HIF-1α-dependent glycolysis and VEGF expression in non-small cell lung cancer cells[J]. Molecular and cellular biochemistry, 2017, 428(1-2): 171-178.

References:

Eolbergrová J, MacMillan V, Siesjö B K. The effect of moderate and marked hypercapnia upon the energy state and upon the cytoplasmic NADH/NAD⁺ ratio of the rat brain[J]. Journal of neurochemistry, 1972, 19(11): 2497-2505.

Related Products:

BC0740/BC0745 Hexokinase(HK) Activity Assay Kit
BC0540/BC0545 Pyruvate Kinase(PK) Activity Assay Kit

BC0530/BC0535 Phosphofructokinase(PFK) Activity Assay Kit

Technical Specifications:

The detection limit:0.0771 μmol/mL The linear range: 0.078-5 μmol/mL