

DPPH Free Radical Scavenging Capacity Assay

Kit

Note: Take two or three different samples for prediction before test.

Operation Equipment: Spectrophotometer

Catalog Number: BC4750

Size: 50T/24S

Components:

Extract Solution: Liquid 50mL×1. Storage at 4°C.

Reagent I: Absolute ethanol 60mL×1. **Self-provided reagent**

Reagent II: Powder×1. Storage at 4°C. (0.6 mL EP tube placed in 8 mL reagent bottle). Add 6.08 mL of reagent I before use to shake and dissolve. Unused reagents can be stored at -20°C for 1 month. It is recommended to store them separately;

According to the required amount of the test sample to prepare **Working solution:** reagent II: reagent I (V: V) =4:21. The unused working solution can be stored at 4°C for a week; The working fluid needs to be temporarily prepared before use.

Reagent III: Powder (VC)×1, Storage at 4°C. Add 1mL extract solution before use to prepare 10mg/mL positive control tube. The unused reagent can be stored at 4°C for 2 weeks.

Product Description

DPPH free radical is a very stable nitrogen-centered free radical. It is one of the important indicators of the sample's antioxidant capacity and is widely used in the research of antioxidant foods, health products and pharmaceuticals.

The DPPH radical has a single electron, and its alcohol solution is purple, with strong absorption at 515 nm. When an antioxidant present, DPPH free radicals are cleared, the solution color becomes lighter, and the absorbance at 515 nm decreases. Within a certain range, the change in absorbance is directly proportional to the degree of free radical removal. In this kit, the ability of the sample to remove DPPH free radicals is reflected by the degree of decrease in absorbance.

Reagents and Equipment Required but Not Provided.

Spectrophotometer, water bath, centrifuge, 1mL glass cuvette, motor/grinder, absolute ethanol, oven, 30-50 mesh sieve and distilled water.

Procedure

I. Sample Preparation

1. Tissue samples: dry fresh samples in a 60°C oven to constant weight, grind them in a mortar (or grinder), and pass a 30-50 mesh sieve. Weigh about 0.05 g sample, add 1 mL of extraction solution, and leaching for 30 min at 40°C water bath. Centrifuge at 10,000 rpm at room temperature for 10 min. Take the

supernatant and place it on ice for testing.

2. Serum, juice or other liquid samples: Pipette 100 μ L of the sample solution into 900 μ L of extraction solution, vortex and mix well, centrifuge at room temperature at 10,000 rpm for 10 min, take the supernatant, and place it on ice for testing.

3. Extract (or drug) can be prepared in a certain concentration, such as 5 mg/mL.

Note: The ability of different samples to remove DPPH free radicals may vary greatly. In order to ensure the accuracy of the experimental results, the samples should be adjusted appropriately according to the results of the pre-experiment (if the removal rate is greater than 90%, it is recommended to dilute the extracted samples with the extraction solution ; The removal rate is less than 5%, it is recommended to increase the quality of the dried sample or the volume of the liquid sample for extraction).

II. Determination Procedure

1. Preheat spectrophotometer for 30 min, adjust the wavelength to 515 nm, set the counter to zero with ethanol.

2. Preparation of positive control tube: If a linear relationship is required, it is recommended that the 10 mg/mL vitamin C solution be prepared into 0.3, 0.25, 0.125, 0.0625, 0.03125, 0.015625 mg/mL with extract solution for use; if 100% clearance rate is required, it is recommended to prepare 0.3 mg/mL vitamin C solution (or greater) with extraction solution.

3. Sample list: add the following reagents to 1.5 mL EP tube.

reagent (μ L)	Blank Tube (A _B)	Test Tube (A _T)	Control Tube (A _C)	Positive control tube (A _P)
Supernatant	-	25	25	-
Reagent III	-	-	-	25
Extract solution	25	-	-	-
Reagent I	-	-	975	-
Working solution	975	975	-	975

After fully mixing, leave it in dark for 30 min at room temperature. Determine the absorbance at 515 nm. The absorbance values of blank tube, control tube, positive control tube and test tube are recorded as A_B, A_C, A_P and A_T respectively. The blank tube only need to be measured 1-2 times.

III. Calculations

1. Formula for the Free Radical Scavenging Rate of the Positive Control:

$$\text{DPPH free radical scavenging rate } D_{VC}\% = [(A_B - A_P) \div A_B] \times 100\%$$

2. Formula for the Free Radical Scavenging Rate of Sample:

$$\text{DPPH free radical scavenging rate } D_{VC}\% = [(A_B - (A_T - A_C)) \div A_B] \times 100\%$$

Note:

1. The DPPH free radical scavenging ability of different samples may vary greatly. If you

want to compare the DPPH free radical scavenging ability of different samples, it is recommended to add the same

amount of samples to the same batch of samples: Add liquid samples such as red wine, tissue homogenate,

and juice to the same volume; The extract (or drug) is formulated to the same concentration.

During the comparison, the sample is adjusted appropriately according to the results of the pre-experiment, and the scavenging rate of the same concentration (same dilution factor) is compared.

2. Samples are recommended to be tested on the day of extraction.

Experimental Examples:

1. Take 0.05g of Leonurus artemisia leaves and add 1mL extract for sample processing, centrifuge to take the supernatant and operate according to the determination steps. It is measured that $A_b=1.037$, $A_c=0.088$, $A_t=0.228$, according to the calculation formula:

DPPH Free Radical Scavenging Capacity $D\% = \frac{[A_b - (A_t - A_c)]}{A_b} \times 100\% = 86.5\%$.

2. Take 100 μ L of red wine and add 900 μ L of extraction solution for sample processing, centrifuge to take the supernatant and operate according to the determination steps. $A_b=1.037$, $A_c=0.012$, $A_t=0.760$, according to the calculation formula:

DPPH Free Radical Scavenging Capacity $D\% = \frac{[A_b - (A_t - A_c)]}{A_b} \times 100\% = 27.9\%$.

3. Take 0.05g of wolfberry and add 1mL extract for sample processing, centrifuge to take the supernatant and dilute it 20 times and follow the determination steps. It is determined that $A_b=1.037$, $A_c=0.005$, $A_t=0.829$, according to the calculation formula:

DPPH Free Radical Scavenging Capacity $D\% = \frac{[A_b - (A_t - A_c)]}{A_b} \times 100\% = 20.5\%$.

Related Products:

BC1310/BC1315 Total Antioxidant Capacity(T-AOC) Assay Kit

BC1320/BC1325 Hydroxyl Free Radical Scavenging Capacity Assay Kit