

# Ferric chelate reductase Activity Assay Kit

**Detection Equipment:** Spectrophotometer

**Catalog Number:** BC5910

**Size:** 50T/48S

**Components:** Please carefully check the volume of the reagent and the volume in the bottle before use. If you have any questions, please contact Solarbio staff in time.

Reagent name	Size	Preservation condition
Extract	Liquid 60 mL×1	2-8°C storage
Reagent I	Liquid 15 mL×1	2-8°C storage
Reagent II	Liquid 15 mL×1	2-8°C storage
Reagent III	Liquid 15 mL×1	2-8°C storage
Standard	Powder ×1	2-8°C storage

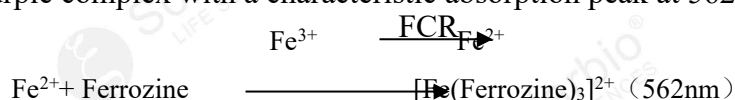
**Solution preparation :**

1. Color development solution: According to the sample quantity before use, according to the ratio of Reagent I: Reagent II: Reagent III =250μL: 250μL: 250μL (750μL, 1T), the color development solution was prepared, mixed thoroughly, and now used;
2. Standard: Add 0.71mL distilled water and 10μL concentrated sulfuric acid to prepare 50μmol/mL Fe<sup>2+</sup> Standard solution before use. The dissolved Standard can be storage for 2 weeks at 2-8°C.
3. Preparation of 62.5nmol/mL Standard solution: 2.5μmol/mL Standard solution was prepared by mixing 50μL of 50μmol/mL Fe<sup>2+</sup> Standard solution and 950μL of distilled water before use. Then 25μL 2.5μmol/mL (2500nmol/mL) and 975μL distilled water were mixed to prepare 62.5nmol/mL Standard solution for later use.

## Description:

Dicots and non-gramineous monocots use efficient activation and uptake mechanisms of iron chelation reduction reactions to obtain iron from soil. Ferric iron can be absorbed and utilized by plants only after it is reduced to divalent iron. When Fe(III) -chelate is sufficient, the trivalent Fe oxidoreductases in plant roots reduce Fe from Fe(III) -chelate and transport the reduced Fe<sup>2+</sup> through the plasma membrane into the root cells.

Ferric chelate reductase (FCR, EC1.16.1.7) catalyzes the reduction of Fe<sup>3+</sup> to Fe<sup>2+</sup>. Fe<sup>2+</sup> and phenoxazine form a purple complex with a characteristic absorption peak at 562nm



**Note:** Before the experiment, it is recommended to select 2-3 sample with large expected differences for pre-experiment. If the absorbance value of the sample is not within the measurement range, it is recommended to dilute or increase the sample size for detection.

## Reagents and Equipment Required but Not Provided:

Spectrophotometer, cryogenic centrifuge, analytical balance, 1mL glass cuvette, adjustable pipetting gun, mortar/homogenizer, concentrated sulfuric acid (95%-99% AR), ice, and distilled

water.

### Protocol:

#### I. Sample processing (The sample size to be tested can be appropriately adjusted, and the specific proportion can be referred to the literature)

- 1. Tissue sample:** Add the extract solution according to the ratio of mass (g) : extraction liquid volume (mL) 1:5-10 (it is recommended to weigh 0.1g sample and add 1.0mL extraction solution). After homogenization in an ice bath, centrifuge at 8000g at 4°C for 10min, discard the precipitate, and take the supernatant and place it on ice for testing.
- 2. Liquid sample:** Direct measurement. If the liquid was cloudy, the supernatant was centrifuged for determination.

#### II. Measurement Steps

- The visible spectrophotometer was preheated for more than 30min, the wavelength was adjusted to 562nm, and zero it with distilled water.
- The chromogenic solution was equilibrated to room temperature before use.
- Standard tube determination  
Absorb 250μL of 62.5nmol/mL Fe<sup>2+</sup> standard solution, add 750μL of color development agent, thoroughly mix, measure the absorbance at 562nm, recorded as standard A, at this time, the final concentration of Fe<sup>2+</sup> is 15.625nmol/mL, the standard tube only needs to do 1-2 times.
- Blank tube test  
Absorb 250μL of distilled water, add 750μL of color development agent, mix thoroughly, measure the absorbance at 562nm, recorded as A blank, calculate  $\Delta A_{\text{standard}} = A_{\text{standard}} - A_{\text{blank}}$ , blank tube only need to do 1-2 times.
- Operation table: (the following reagents in 1mL glass cuvette)

Reagent name (μL)	Test tube
Sample	250
Color developing solution	750

Immediately mix thoroughly in 1mL glass cuvette, the absorbance value A1 at 10s was measured at 562nm, and the reaction time was 30min at room temperature. The absorbance value A2 at 10s was measured at 30min, and the absorbance value A1 at 10s at 562nm and A2 after 30min were recorded.  $\Delta A = A2 - A1$  was calculated.

#### III. Calculations

##### 1. According to the sample protein concentration:

Definition of unit: 1nmol Fe<sup>2+</sup> produced per minute per mg of tissue protein was defined as one unit of enzyme activity.

$$\text{FCR activity (U/mg prot)} = \Delta A \times C_{\text{standard}} \div \Delta A_{\text{standard}} \times V_{\text{total}} \div (C_{\text{pr}} \times V_{\text{sample}}) \div T = 2.083 \times \Delta A \div \Delta A_{\text{standard}} \div C_{\text{pr}}$$

##### 2. According to sample quality:

Definition of unit: 1nmol Fe<sup>2+</sup> produced per minute per g of tissue was defined as one unit of

enzyme activity.

$$\text{FCR activity (U/g)} = \Delta A \times C_{\text{standard}} \div \Delta A_{\text{standard}} \times V_{\text{total reaction}} \div (W \div V_{\text{extraction}} \times V_{\text{sample}}) \div T$$

$$= 2.083 \times \Delta A \div \Delta A_{\text{standard}} \div W$$

$C_{\text{standard}}$ :  $\text{Fe}^{2+}$  standard final concentration, 15.625nmol/mL;

$V_{\text{total reaction}}$ : total volume of reaction system, 1mL;

$C_{\text{pr}}$ : supernatant protein concentration, mg/mL;

$V$ : Add the volume of supernatant liquid in the reaction system, 250 $\mu$ L=0.25mL;

$V_{\text{extraction}}$ : add extraction liquid volume, 1mL;

$T$ : reaction time, 30min;

$W$ : sample quality, g.

#### Note:

1. If  $\Delta A$  is less than 0.010 or the absorbance value of the measuring tube is close to the blank tube, the sample size can be increased or the reaction time can be prolonged before the measurement; If  $\Delta A$  is greater than 1 or  $A_1$  is greater than 1, it is recommended that the sample supernatant be appropriately diluted with the extract before performing the assay. Note that the calculation formula is modified synchronously.

#### Experimental example:

1. Weigh 0.1214g of cherry tomatoes, add the extract to the ice bath homogenate, and follow the determination steps. Calculated with 1mL glass cuvette  $\Delta A = A_2 - A_1 = 0.921 - 0.082 = 0.839$ ,  $\Delta A_{\text{standard}} = A_{\text{standard}} - A_{\text{blank}} = 0.723 - 0.044 = 0.679$ , and put them into the formula to calculate:

$$\text{FCR activity (U/g mass)} = 2.083 \times \Delta A \div \Delta A_{\text{standard}} \div W = 21.020 \text{ U/g mass}$$

2. Weigh 0.1002g Minnan leaves, add the extract for ice bath homogenization, and follow the determination steps. Calculated with 1mL glass cuvette  $\Delta A = A_2 - A_1 = 0.203 - 0.088 = 0.115$ ,  $\Delta A_{\text{standard}} = A_{\text{standard}} - A_{\text{blank}} = 0.723 - 0.044 = 0.679$ , and put them into the formula to calculate:

$$\text{FCR activity (U/g mass)} = 2.083 \times \Delta A \div \Delta A_{\text{standard}} \div W = 3.521 \text{ U/g mass}$$

#### References:

[1] Shabnam N , Kim M , Kim H .Iron (III) oxide nanoparticles alleviate arsenic induced stunting in *Vigna radiata*[J].Ecotoxicology and Environmental Safety, 2019, 183(Nov.).

[2] Kabir A H , Akther M S , Skalicky M ,et al.Downregulation of Zn-transporters along with Fe and redox imbalance causes growth and photosynthetic disturbance in Zn-deficient tomato[J].Scientific Reports, 2021, 11(1).

[3] Ojeda M , Schaffer B , Davies F S .Root and Leaf Ferric Chelate Reductase Activity in Pond Apple and Soursop[J].Journal of Plant Nutrition, 2005, 27(8):1381-1393.

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